

# NATIONAL CENTER FOR COMPETENCY TESTING (NCCT) PRACTICE EXAM (SAMPLE) STUDY GUIDE



## SAMPLE STUDY GUIDE WITH LIMITED QUESTIONS

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# Introduction

Preparing for a certification exam can feel overwhelming, but with the right tools, it becomes an opportunity to build confidence, sharpen your skills, and move one step closer to your goals. At Examzify, we believe that effective exam preparation isn't just about memorization, it's about understanding the material, identifying knowledge gaps, and building the test-taking strategies that lead to success.

This guide was designed to help you do exactly that.

Whether you're preparing for a licensing exam, professional certification, or entry-level qualification, this book offers structured practice to reinforce key concepts. You'll find a wide range of multiple-choice questions, each followed by clear explanations to help you understand not just the right answer, but why it's correct.

The content in this guide is based on real-world exam objectives and aligned with the types of questions and topics commonly found on official tests. It's ideal for learners who want to:

- Practice answering questions under realistic conditions,
- Improve accuracy and speed,
- Review explanations to strengthen weak areas, and
- Approach the exam with greater confidence.

We recommend using this book not as a stand-alone study tool, but alongside other resources like flashcards, textbooks, or hands-on training. For best results, we recommend working through each question, reflecting on the explanation provided, and revisiting the topics that challenge you most.

Remember: successful test preparation isn't about getting every question right the first time, it's about learning from your mistakes and improving over time. Stay focused, trust the process, and know that every page you turn brings you closer to success.

Let's begin.

# How to Use This Guide

This guide is designed to help you study more effectively and approach your exam with confidence. Whether you're reviewing for the first time or doing a final refresh, here's how to get the most out of your Examzify study guide:

## 1. Start with a Diagnostic Review

Skim through the questions to get a sense of what you know and what you need to focus on. Your goal is to identify knowledge gaps early.

## 2. Study in Short, Focused Sessions

Break your study time into manageable blocks (e.g. 30 – 45 minutes). Review a handful of questions, reflect on the explanations.

## 3. Learn from the Explanations

After answering a question, always read the explanation, even if you got it right. It reinforces key points, corrects misunderstandings, and teaches subtle distinctions between similar answers.

## 4. Track Your Progress

Use bookmarks or notes (if reading digitally) to mark difficult questions. Revisit these regularly and track improvements over time.

## 5. Simulate the Real Exam

Once you're comfortable, try taking a full set of questions without pausing. Set a timer and simulate test-day conditions to build confidence and time management skills.

## 6. Repeat and Review

Don't just study once, repetition builds retention. Re-attempt questions after a few days and revisit explanations to reinforce learning. Pair this guide with other Examzify tools like flashcards, and digital practice tests to strengthen your preparation across formats.

There's no single right way to study, but consistent, thoughtful effort always wins. Use this guide flexibly, adapt the tips above to fit your pace and learning style. You've got this!

## Questions

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- 1. What is the proper method to clean up a broken glass tube?**
  - A. Use a paper towel to pick up the pieces and place them in a trash can
  - B. Place the pieces on paper and carry to red biohazard bag
  - C. Use a broom and dust pan to sweep up the glass and fragments and place in a sharps container
  - D. Use a wet towel and wipe over the area so that minute fragments of glass may be picked up
- 2. How long should the heel be warmed before capillary blood collection?**
  - A. 1-2 minutes
  - B. 3-5 minutes
  - C. 6-8 minutes
  - D. 10 minutes
- 3. What is the CLSI recommended micro-collection order of draw for a BMP, CBC and bilirubin?**
  - A. Lavender, green, red
  - B. Green, lavender, red
  - C. Lavender, red, green
  - D. Red, green, lavender
- 4. Which QC measure is within the phlebotomist's scope of practice?**
  - A. Calibrating the chemistry analyzer
  - B. Daily maintenance on the laboratory hematology instruments
  - C. Checking the expiration dates of venipuncture supplies
  - D. None of the above
- 5. Which item is NOT listed as PPE in the described isolation precautions scenario with gown, gloves, and mask?**
  - A. Gown
  - B. Goggles
  - C. Gloves
  - D. Mask

- 6. Immediately before removing the needle, which action should the phlebotomist take?**
- A. Activate needle safety mechanism
  - B. Release the tourniquet
  - C. Gently invert the evacuated tubes
  - D. Apply a gauze pressure dressing
- 7. Which statement best describes why red-top tubes are used for serum testing?**
- A. They contain anticoagulants
  - B. They contain clot activators
  - C. They contain no additives
  - D. They are permanently sterile
- 8. A woman who becomes sensitized to the Rh factor can produce antibodies against which antigen?**
- A. Anti-A
  - B. Anti-B
  - C. Rh antigen
  - D. A and B antigens
- 9. In a PT and CBC collection, which sequence of evacuated tube colors is correct?**
- A. Lavender, light blue
  - B. Light blue, lavender
  - C. Light blue, red
  - D. Red, light blue
- 10. Which outcome is most directly addressed by keeping the stopper in place during processing?**
- A. Contamination
  - B. Analyte dilution
  - C. Decrease in pH
  - D. Temperature change

## **Answers**

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1. C
2. B
3. C
4. C
5. B
6. B
7. B
8. A
9. B
10. A

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## **Explanations**

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### 1. What is the proper method to clean up a broken glass tube?

- A. Use a paper towel to pick up the pieces and place them in a trash can
- B. Place the pieces on paper and carry to red biohazard bag
- C. Use a broom and dust pan to sweep up the glass and fragments and place in a sharps container**
- D. Use a wet towel and wipe over the area so that minute fragments of glass may be picked up

*Broken glass is a sharp hazard, so the goal is to remove it safely with the right tools and proper disposal. Use a broom and dustpan to sweep up the fragments so you can collect all pieces without risking cuts. Then place the collected glass in a sharps container—the puncture resistant, clearly labeled container used for sharp waste. This protects you and others and ensures the waste is handled correctly during disposal. Disposing in a regular trash bag or biohazard bag isn't appropriate for clean glass, and trying to wipe with a wet towel or picking up with a paper towel can leave tiny shards behind or fail to contain them. Vacuuming is usually avoided because it can spread shards or damage the equipment.*

### 2. How long should the heel be warmed before capillary blood collection?

- A. 1-2 minutes
- B. 3-5 minutes**
- C. 6-8 minutes
- D. 10 minutes

*Warming the heel increases blood flow to the area by dilating the capillaries, which makes capillary blood collection easier and helps you obtain an adequate sample with less discomfort. The recommended duration is about three to five minutes. Warming for a short time may not boost perfusion enough, making it harder to obtain a good blood drop. Warming for much longer than needed can irritate or damage the skin and potentially affect the sample. Use a clean, warm, moist pad at a safe temperature and apply it for roughly three to five minutes before puncturing.*

### 3. What is the CLSI recommended micro-collection order of draw for a BMP, CBC and bilirubin?

- A. Lavender, green, red
- B. Green, lavender, red
- C. Lavender, red, green**
- D. Red, green, lavender

*The idea being tested is how to order draws so additives in one tube don't contaminate the others. Lavender tubes contain EDTA for the CBC and protect cell integrity, red tubes are serum tubes with no anticoagulant, and green tubes contain heparin for plasma chemistry like a BMP. Following the CLSI micro-collection guideline, drawing CBC first in the EDTA tube, then bilirubin in a red serum tube, and finally BMP in a green heparin tube minimizes the chance that EDTA or other additives carry over into the chemistry samples. If the chemistry tubes were drawn sooner, EDTA or heparin could contaminate the serum or plasma samples and distort results, especially in chemistry measurements and calcium-related assays.*

**4. Which QC measure is within the phlebotomist's scope of practice?**

- A. Calibrating the chemistry analyzer
- B. Daily maintenance on the laboratory hematology instruments
- C. Checking the expiration dates of venipuncture supplies**
- D. None of the above

*Phlebotomists focus on the pre-analytical phase to protect specimen quality. Checking expiration dates on venipuncture supplies fits this scope because it directly ensures the materials used for collection are valid and won't compromise the specimen or test results. Using expired tubes, additives, or supplies can affect specimen integrity and lead to unreliable results, which is something a phlebotomist is responsible for preventing. Calibrating a chemistry analyzer and performing daily maintenance on hematology instruments are instrument QC tasks that require specialized training and certification in laboratory technology, and they are outside the phlebotomist's typical duties. Therefore, the task within the phlebotomist's scope is checking expiration dates.*

**5. Which item is NOT listed as PPE in the described isolation precautions scenario with gown, gloves, and mask?**

- A. Gown
- B. Goggles**
- C. Gloves
- D. Mask

*The key idea here is what PPE is considered for a given isolation scenario. Gown and gloves protect the skin and clothing, while a mask protects the mouth and nose from droplets. Eye protection—like goggles or a face shield—is added only when there's a risk of splashes, sprays, or other exposure to the eyes. In the described setup those three items are listed, but eye protection isn't mentioned, so goggles are the one not included in this scenario. If a procedure or situation raises risk to the eyes, eye protection would be added on top of the gown, gloves, and mask.*

**6. Immediately before removing the needle, which action should the phlebotomist take?**

- A. Activate needle safety mechanism
- B. Release the tourniquet**
- C. Gently invert the evacuated tubes
- D. Apply a gauze pressure dressing

*Releasing the tourniquet just before removing the needle is the step that helps control bleeding and prevent a hematoma. When the tourniquet is still on, the vein is engorged and blood is under higher pressure. Pulling the needle out with that pressure can cause more bleeding into surrounding tissue, leading to a hematoma and prolonged bleeding. By releasing the tourniquet, venous pressure drops, making it safer to withdraw the needle and then immediately apply gauze with pressure to achieve hemostasis. Gently inverting evacuated tubes happens during blood collection after tubes are attached, not right before needle withdrawal. Activating the needle safety mechanism is typically handled as part of or after removing the needle. Applying a gauze pressure dressing is done after the needle is removed, not before.*

**7. Which statement best describes why red-top tubes are used for serum testing?**

- A. They contain anticoagulants
- B. They contain clot activators**
- C. They contain no additives
- D. They are permanently sterile

*Red-top tubes used for serum testing are designed to help the blood clot so serum can be separated from the clot. The clot activators inside many of these tubes speed up the coagulation process, making it quicker and more consistent to obtain serum. If a tube contained anticoagulants, the blood wouldn't clot and you'd get plasma instead of serum, which changes the type of specimen and the tests you can run. Some red-top tubes have no additives, but in many modern setups the clot activator is what makes them suitable for efficient serum collection. Being permanently sterile isn't related to why these tubes are used for serum testing.*

**8. A woman who becomes sensitized to the Rh factor can produce antibodies against which antigen?**

- A. Anti-A**
- B. Anti-B
- C. Rh antigen
- D. A and B antigens

*The key idea is that Rh sensitization prompts the formation of antibodies against the Rh antigen, specifically the D antigen. When a person who is Rh-negative is exposed to Rh-positive red blood cells—such as during pregnancy with an Rh-positive fetus or from a transfusion—they can develop anti-D antibodies. These antibodies target the Rh antigen on future Rh-positive cells. Anti-A and anti-B antibodies are part of the ABO system and arise from exposure to A or B antigens, not from Rh sensitization. So, the antibodies produced in Rh sensitization are directed against the Rh antigen (D).*

**9. In a PT and CBC collection, which sequence of evacuated tube colors is correct?**

- A. Lavender, light blue
- B. Light blue, lavender**
- C. Light blue, red
- D. Red, light blue

*This question tests the correct order of draw when collecting blood for coagulation testing (PT) together with a CBC. The light blue tube contains sodium citrate, an anticoagulant that preserves plasma for coagulation tests by preventing clotting while maintaining the proper blood-to-anticoagulant ratio. Because this ratio can be spoiled by introducing other additives, the light blue tube should be drawn before any tubes with other additives. The lavender tube contains EDTA, which preserves cells for CBC by chelating calcium and preventing clotting in the cells. If EDTA were drawn first or before the light blue tube, EDTA could contaminate the coagulation sample and skew PT results. Therefore, the best sequence is to draw the light blue tube first, followed by the lavender tube. Other sequences risk contamination of the coagulation sample or fail to provide the properly preserved CBC sample.*

**10. Which outcome is most directly addressed by keeping the stopper in place during processing?**

**A. Contamination**

B. Analyte dilution

C. Decrease in pH

D. Temperature change

*Keeping the stopper in place primarily protects the sample from outside contaminants. It acts as a barrier to air, dust, microbes, and other potential intruders, preserving the integrity of the analyte during processing. While an in-place stopper can also help reduce evaporation and spills, the most direct and important effect is preventing contamination, which could otherwise skew results or require re-testing. Temperature changes and pH shifts are influenced by many factors beyond simply sealing the container, so they're not addressed as directly by leaving the stopper in place.*

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## Next Steps

Congratulations on reaching the final section of this guide. You've taken a meaningful step toward passing your certification exam and advancing your career.

As you continue preparing, remember that consistent practice, review, and self-reflection are key to success. Make time to revisit difficult topics, simulate exam conditions, and track your progress along the way.

If you need help, have suggestions, or want to share feedback, we'd love to hear from you. Reach out to our team at [hello@examzify.com](mailto:hello@examzify.com).

Or visit your dedicated course page for more study tools and resources:

<https://ncct.examzify.com>

We wish you the very best on your exam journey. You've got this!

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